

Counterion Analysis

Phosphate as an example for Drug stoichiometry and impurity determination using Capillary Electrophoresis

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2nd Joint CE user Meeting

Outlook

- Pharmaceutical Counterions
- Analysis of anions, organic acids and cations with indirect UV detection
- Analysis of PO_4 as counterion
- Analysis of PO_4 as impurity

Use of CE for pharmaceutical analysis

1. Chiral analysis
2. Impurity (CE-MS)
- 3. Counterions**
4. pKa determination
5. Cleaning control
6. Protein/peptides

HPCE Stockholm 2002

Pharmaceutical Counterions

- Most drugs are weak bases or acids
- Most drugs are produced as a salt
- Regulatory agencies (FDA, European Pharmacopeia...):

....pharmaceutical products should be tested for composition of their identity, strength, quality and purity of the active as well as for the inactive ingredients....

Pharmaceutical Counterions

Counterion quantification needed for
P (purity)

$$P = 100\% - (m/M\% \text{ impurities} + m/M\% \text{ water} + m/M\% \text{ remaining solvents} + m/M\% \text{ inorganic material} + m/M\% \text{ Counterion})$$

Example: Sufentanyl citraat
total MW 578.69 \Rightarrow 386.56 + 192.13

Organic Anions

Acetate C ₂ H ₃ O ₂		
Benzoate (-1) C ₇ H ₅ O ₂		
Benzoate C ₇ H ₆ O ₂		
Citrate (-1) C ₆ H ₇ O ₇		
Citrate (-2) C ₆ H ₆ O ₇		
Citrate (-3) C ₆ H ₅ O ₇		
Citrate C ₆ H ₈ O ₇		
(+/-)-Camphorsulfonate C ₁₀ H ₁₆ O ₄ S	(1R)-(-)-10-Camphorsulfonate	(1S)-(+)-10-Camphorsulfonate
Dibenzoyl-L-tartrate (-1) C ₁₈ H ₁₃ O ₈	Dibenzoyl-D-tartrate (-1)	
Dibenzoyl-L-tartrate (-2) C ₁₈ H ₁₂ O ₈	Dibenzoyl-D-tartrate (-2)	
Dibenzoyl-L-tartrate C ₁₈ H ₁₄ O ₈	Dibenzoyl-DL-tartrate	
Dibenzoyl-L-tartrate C ₁₈ H ₁₄ O ₈	Dibenzoyl-D-tartrate	
Di-p-Toluoyl-D-tartrate C ₂₀ H ₁₈ O ₈	Di-p-Toluoyl-L-tartrate	
Fumarate (-1) C ₄ H ₃ O ₄		
Fumarate (-2) C ₄ H ₂ O ₄		
Fumarate C ₄ H ₄ O ₄ (E)--2-Butenedioic acid	Maleate Z-2-butenedioic acid	
2-Hydroxybenzoate (-1) C ₇ H ₅ O ₃		
Hydroxybutanedioate C ₄ H ₆ O ₅	(R)-Hydroxybutanedioate	(S)-Hydroxybutanedioate
(+/-)-2-Hydroxypropanoate (-1) C ₃ H ₅ O ₃		
(+/-)-2-Hydroxypropanoate C ₃ H ₆ O ₃	(S)-(+)-2-Hydroxypropanoate	

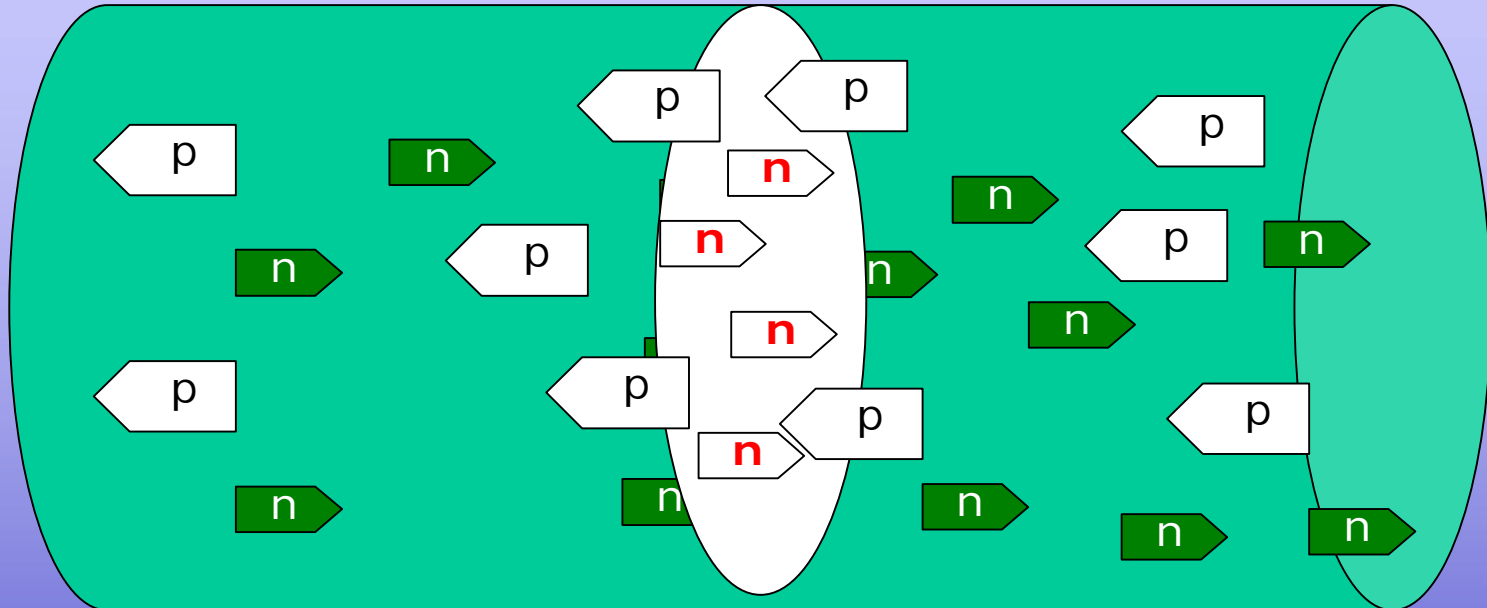
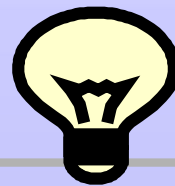
Maleate (-1) C ₄ H ₃ O ₄		
Maleate (-2) C ₄ H ₂ O ₄		
Malonate (-1) C ₃ H ₃ O ₄		
Malonate (-2) C ₃ H ₂ O ₄		
Malonate C ₃ H ₄ O ₄		
(R) Mandelate C ₈ H ₈ O ₃	(S)-mandelate	
Mandelate (-1) C ₈ H ₇ O ₃		
Methaansulfonate (-1) CH ₃ O ₃ S		
Methaansulfonate CH ₄ O ₃ S		
4-Methylbenzenesulfonate C ₇ H ₈ O ₃ S		
Oxalate (-1) C ₂ H ₀ O ₄		
Oxalate (-2) C ₂ O ₄		
Oxalate C ₂ H ₂ O ₄		
Propanoate C ₃ H ₆ O ₂		
Succinate (-1) C ₄ H ₅ O ₄		
Succinate (-2) C ₄ H ₄ O ₄		
Succinate C ₄ H ₆ O ₄		
Tartrate C ₄ H ₆ O ₆	(-)-D-Tartrate	(+)-L-Tartrate
(-)-D-Tartrate (-1) C ₄ H ₅ O ₆	(+)-L-Tartrate (-1)	
(-)-D-Tartrate (-2) C ₄ H ₄ O ₆	(+)-L-Tartrate (-2)	
(+/-)-DL-Tartrate C ₄ H ₆ O ₆		

<i>Inorganic Anions</i>
Nitrate HNO ₃
Phosphate H ₃ PO ₄
Sulfate H ₂ SO ₄
Sulfate (-1) HSO ₄
Sulfate (-2) SO ₄
HCl
HBr


Cations
Ammonium (+1)
Calcium (+2)
Potassium (+1)
Magnesium (+2)
Sodium (+1)

Buffers for Indirect UV detection

- principle
- UV-absorbing probe
- Inverse the EOF
- pH – molarity of the buffer



Buffer:

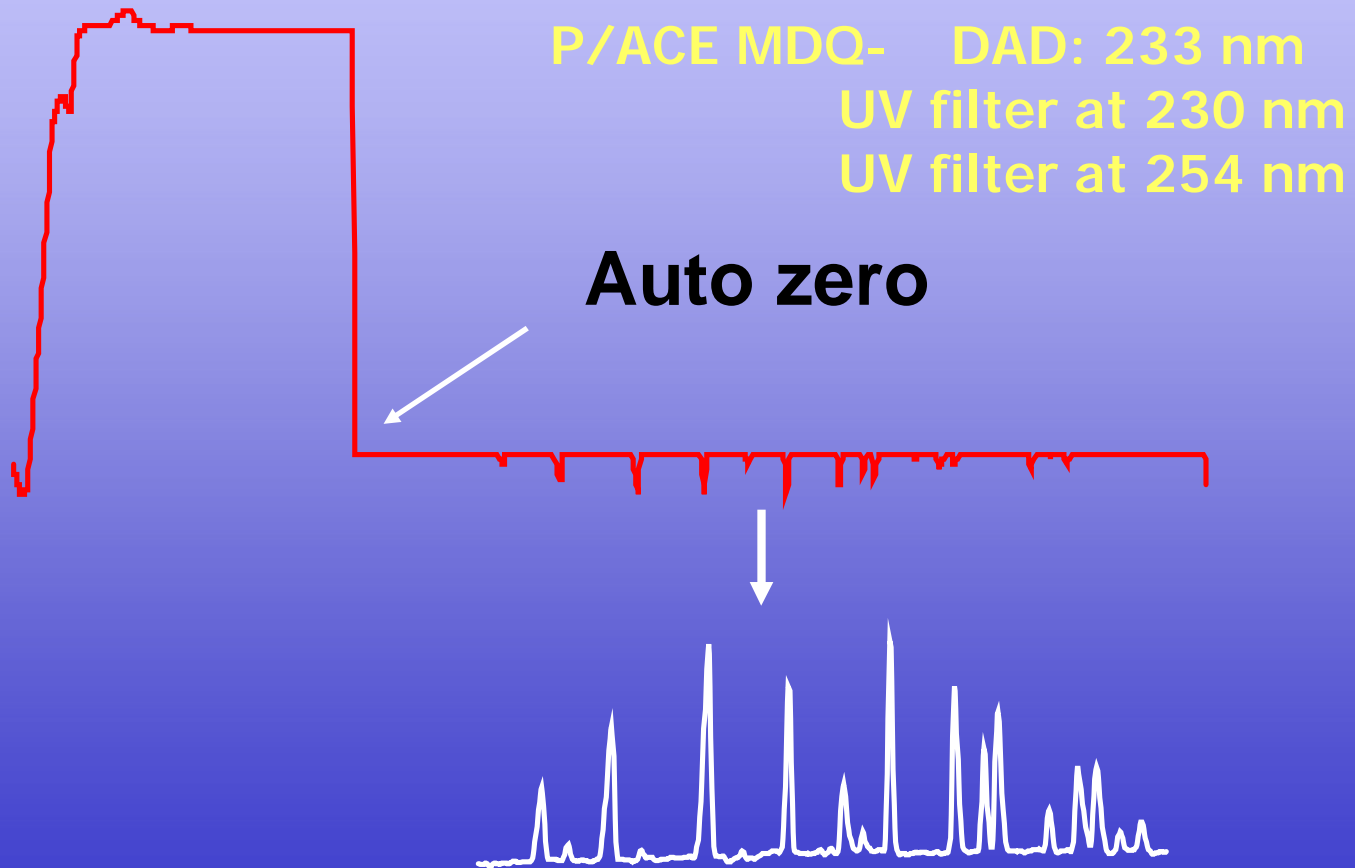
 co-ion chromophore
= *probe*

 counterion

Sample



Detection: indirect-UV



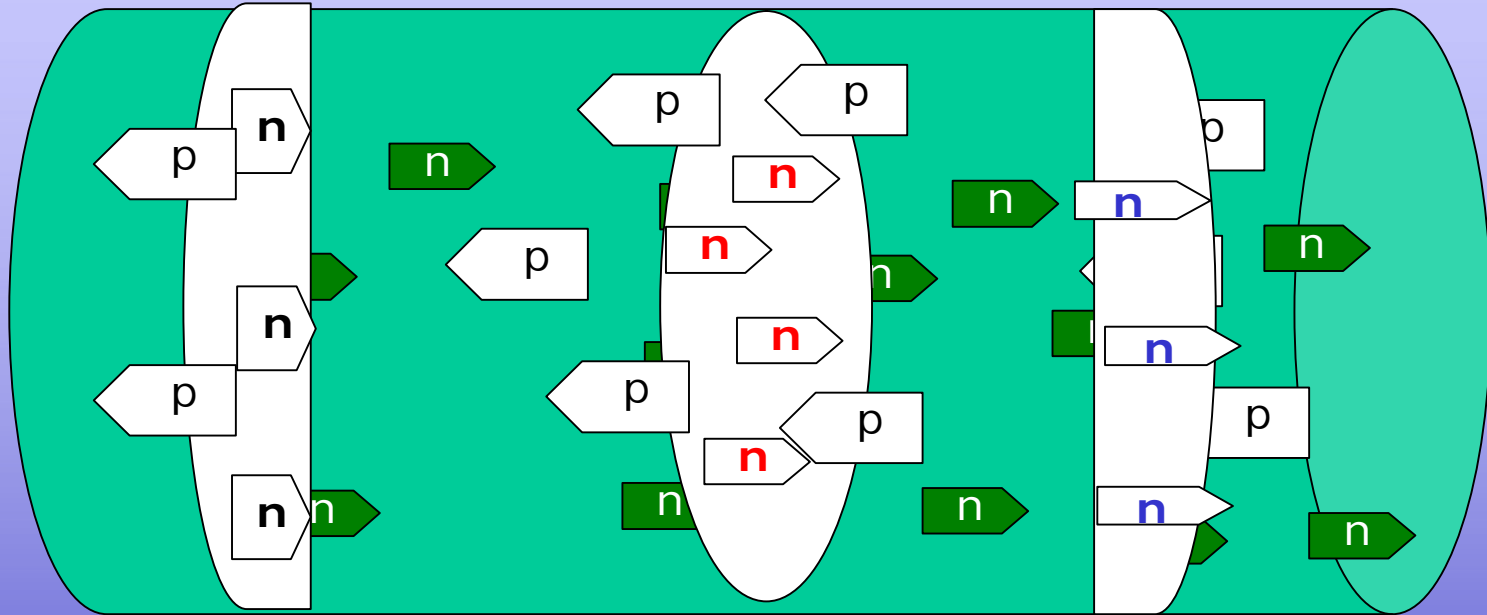
- Transfert Ratio

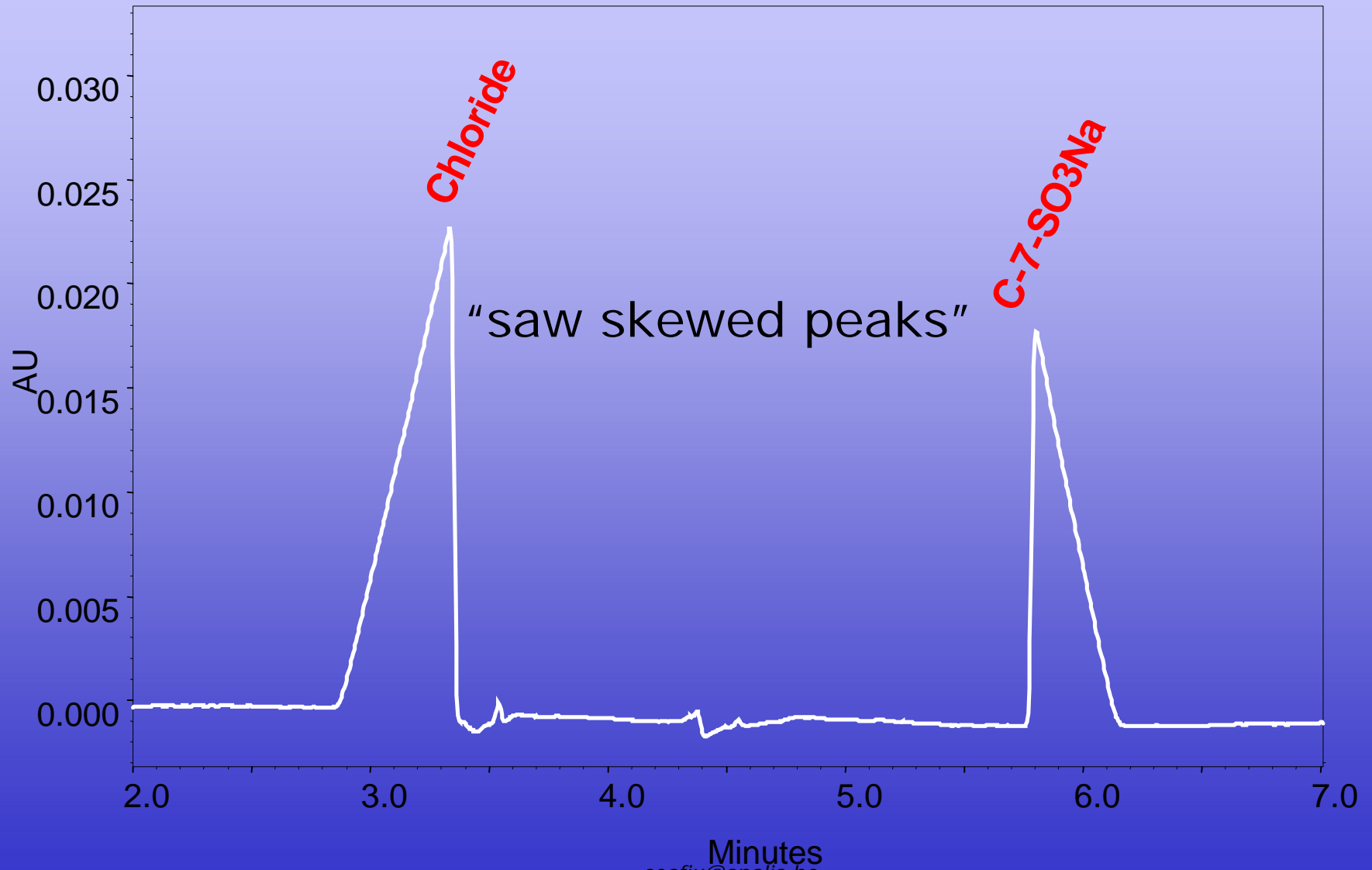
$R = \text{number of moles of the probe displaced by one mole of analyte ion}$

- Kohlrausch Regulating Function(KRF)
= constant

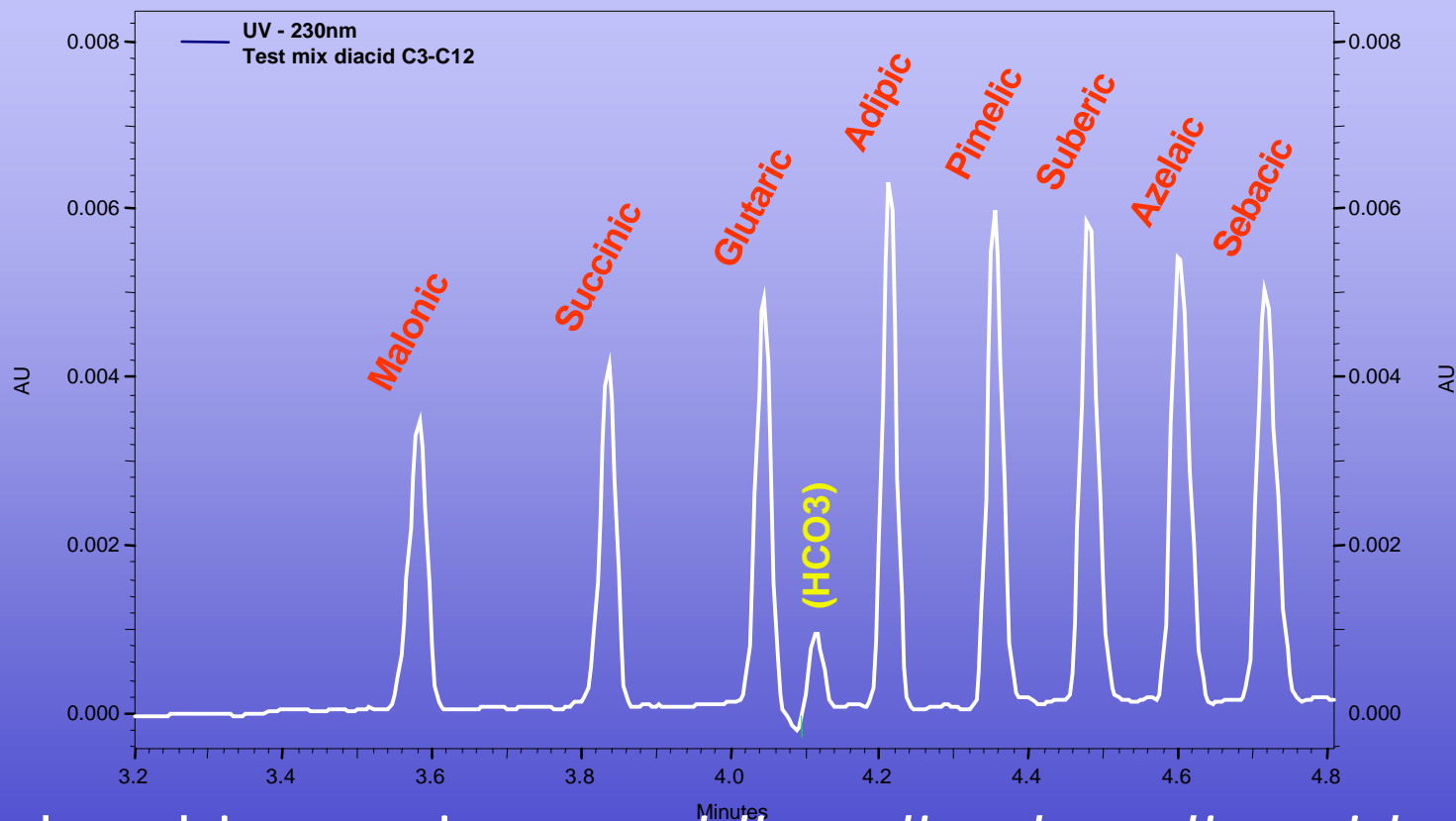
$$? = \sum_i \frac{C_i Z_i}{\mu_i}$$

C = ionic concentration
z = value of the charge
 μ = effective mobility

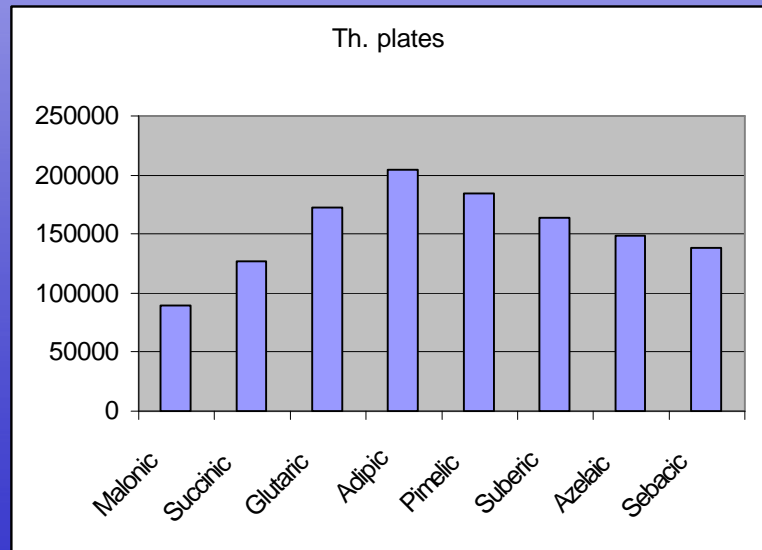
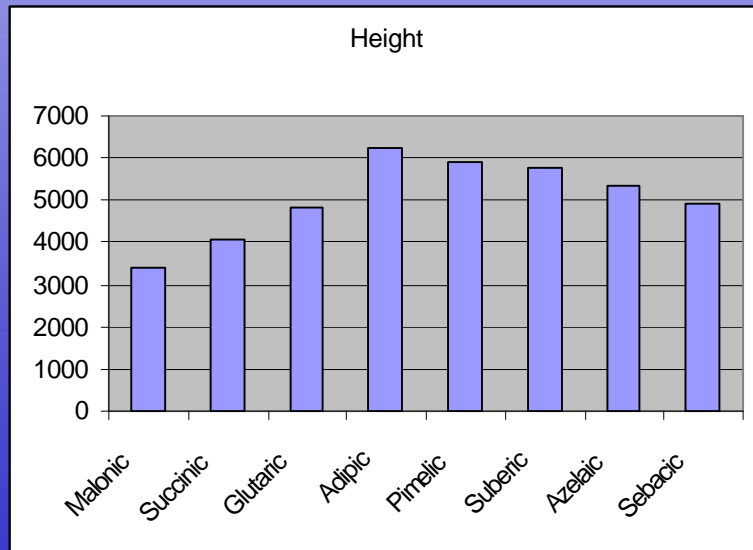
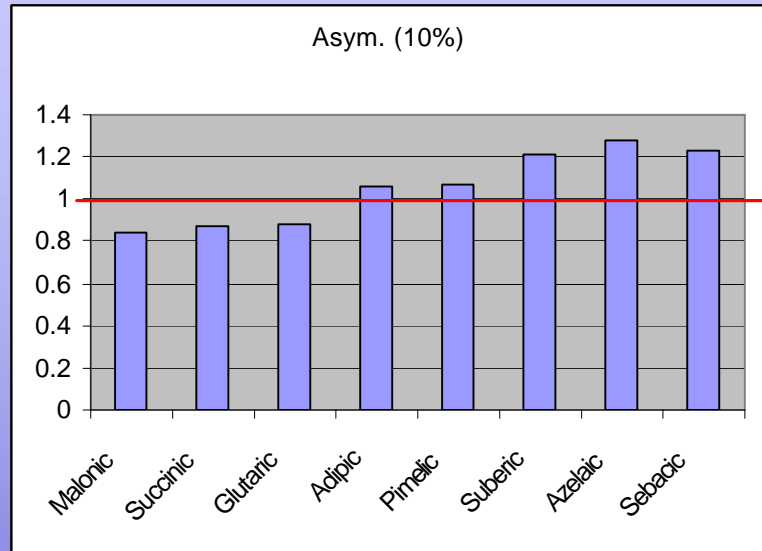
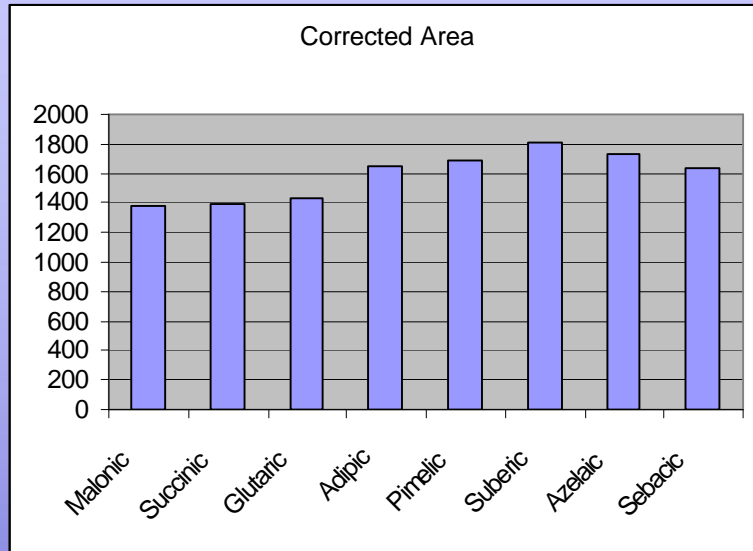




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- absorbing co-ion: *pyridine-dicarboxylic acid*
- pH 8.2



- Limit of detection (C_{LOD})

$$C_{LOD} = \frac{N_{BL}}{R e I}$$

N_{BL} = baseline noise
 e = molar absorptivity of the probe
 I = detection path length
 R = transfer ratio

- Dynamic Reserve (D_r)

$$D_r = \frac{S}{s}$$

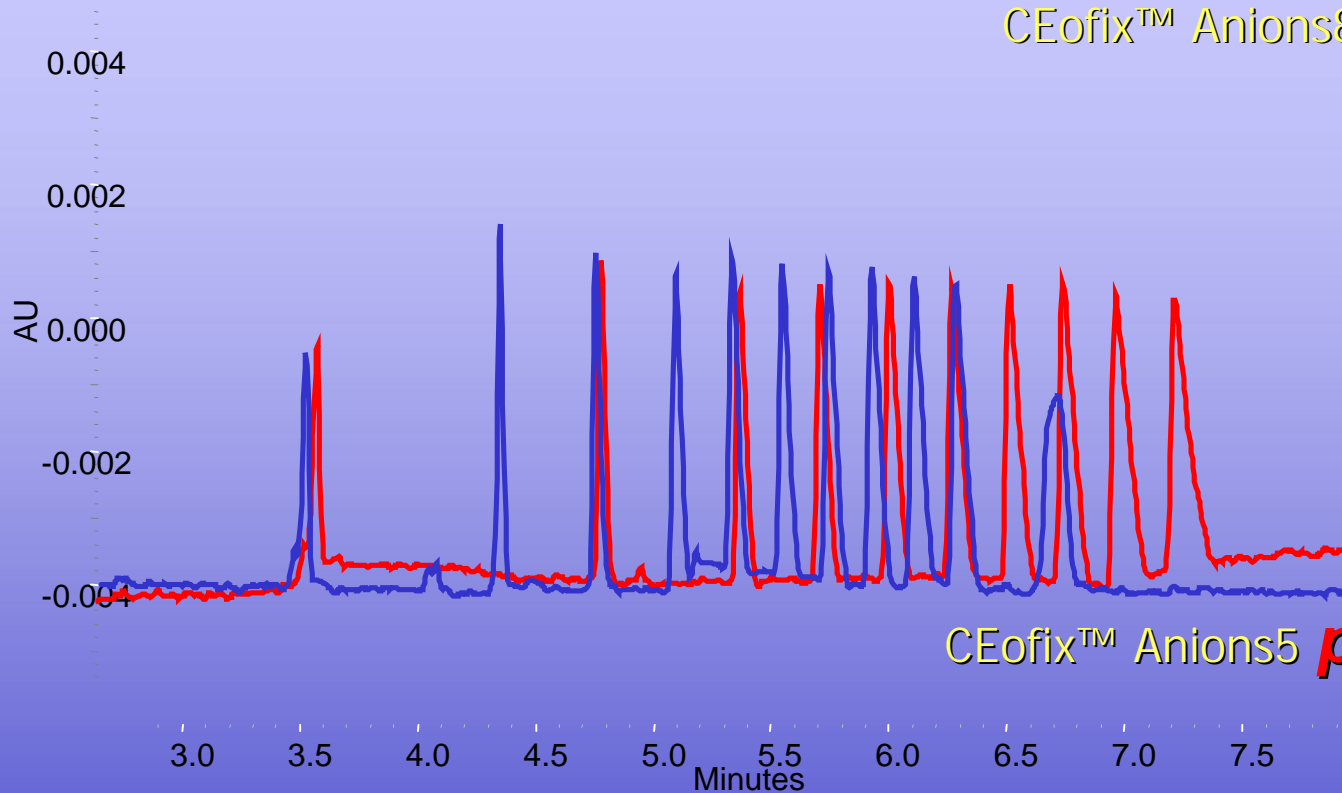
S = signal of detector
 s = standard deviation of the signal

Absorbing Probe

- mobility close to the mobility of the analyte
- co-ion of the buffer: *phthalate*, *pyridinedicarboxylic acid*....
- specific co-ion: *chromate*, *molybdate*...
- high molar absorptivity
- soluble in the buffer

BGE effect of pH

CEofix™ Anions8 **pH 8.2**

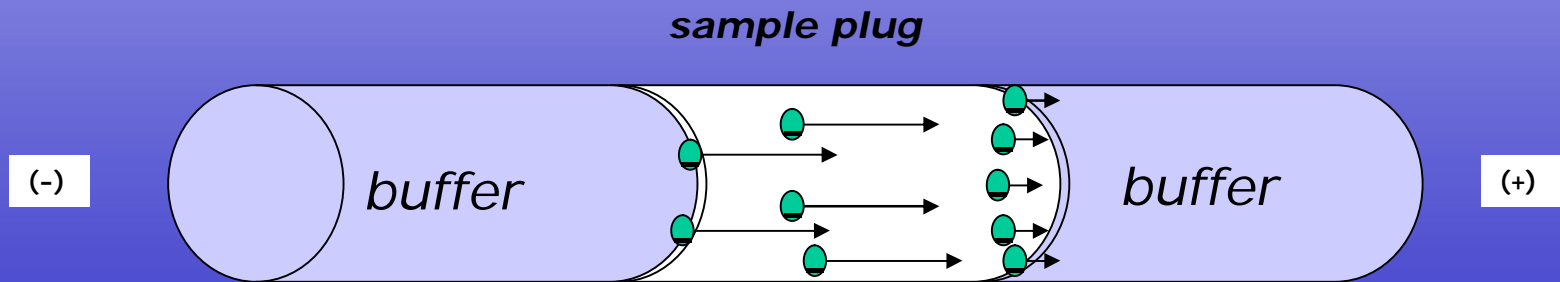


CEofix™ Anions5 **pH 5.6**

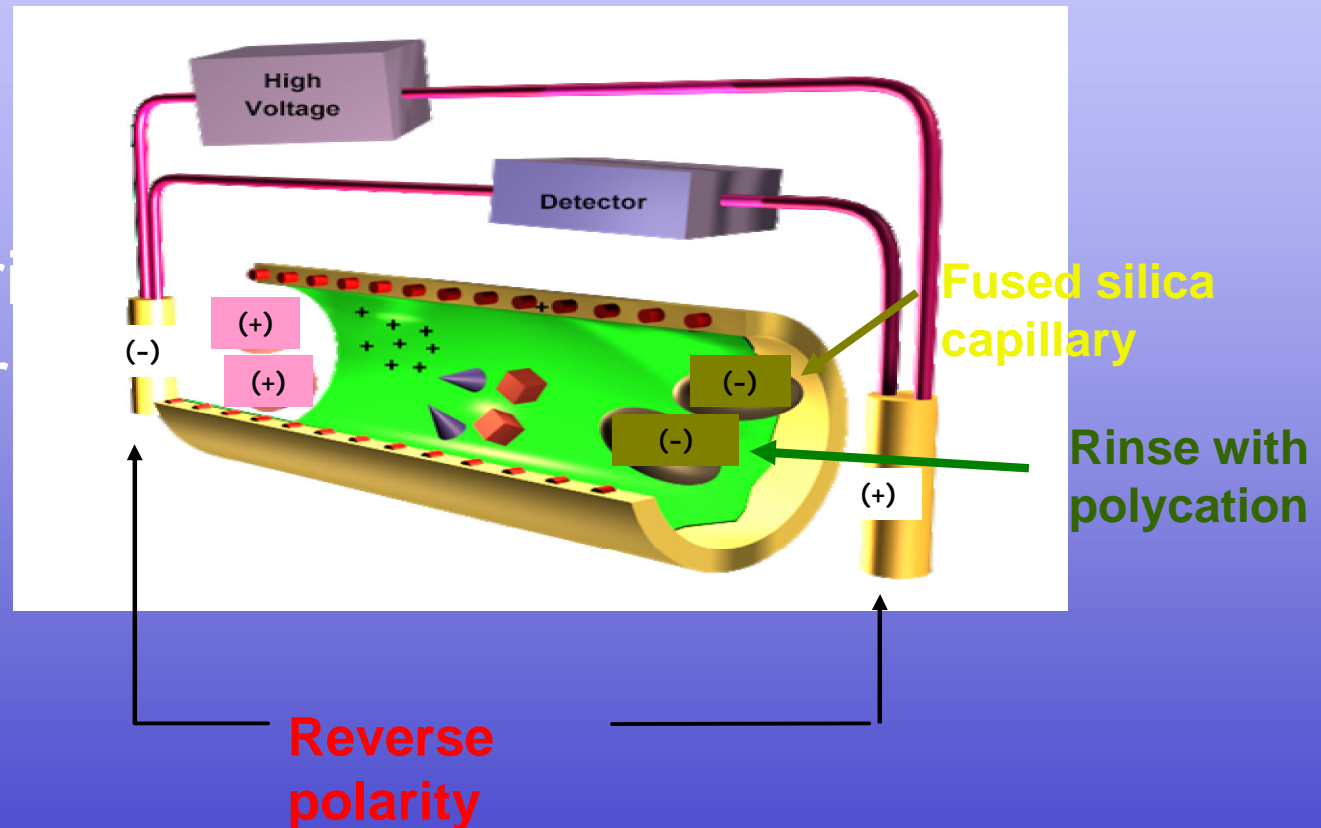
1	Formic 3.74	4	Butyric 4.87	7	Heptanoic 4.89	10	Capric 4.9
2	Acetic 4.76	5	Valeric 4.81	8	Caprylic 4.89	11	-
3	Propionic 4.87	6	Caproic 4.80	9	Pelargonic 4.96	12	Lauric 4.9

Background Electrolyte

- BGE: pH, ionic strength, viscosity
 - buffer
 - pKa of the analyte
 - 10 x higher as the ionic strength of sample: FASS



- reverse polarity
- EOF modifier (surfactant)



EOF flow modifiers

- To inverse the EOF flow
- Cationic surfactants:
CTAB, CTAH, TTAB...
- Polyanion:
Polybrene...

Buffers for Indirect UV detection which system to chose?

- EOF-flow modifier ?
- Buffer ?
- Absorbing Co-ion with matched mobility ?
- What about system peaks?

In most cases a validated buffer system will provide a practical solution!

Instrument considerations

- Capillary
 - length (60 cm total length)
 - diameter (50 – 75 μ)
- Instrument
 - temperature
 - pH of the buffer
 - viscosity of the buffer
 - voltage
- Method

	Time (min)	Event	Value	Duration	Inlet vial	Outlet vial	Summary	
1		Rinse - Pressure	20.0 psi	0.50 min	BI:B1	BO:A1	forward	Initiator
2		Rinse - Pressure	20.0 psi	0.50 min	BI:C1	BO:A1	forward	Accelerator
3		Inject - Pressure	0.5 psi	8.0 sec	SI:A1	BO:C1	No override, forward, In vial inc 1	Sample-H2O
4		Inject - Pressure	0.1 psi	10.0 sec	BI:D1	BO:C1	No override, forward	H2O-H2O
5	0.00	Separate - Voltage	30.0 KV	8.00 min	BI:E1	BO:B1	1.00 Min ramp, reverse polarity	Buffer-Buffer
6	1.25	Autozero						
7	8.00	Stop data						
8	8.00	Rinse - Pressure	20.0 psi	0.50 min	BI:A1	BO:A1	forward	NaOH 0.1M-Waste
9	8.50	Rinse - Pressure	20.0 psi	0.50 min	BI:F1	BO:A1	forward	H2O-Waste
10	9.00	End						

- rinse step
- H₂O post-injection ? H₂O rinse step
- buffer rinse ? buffer separation step
- 1 min. ramping time
- increment every 20 analysis

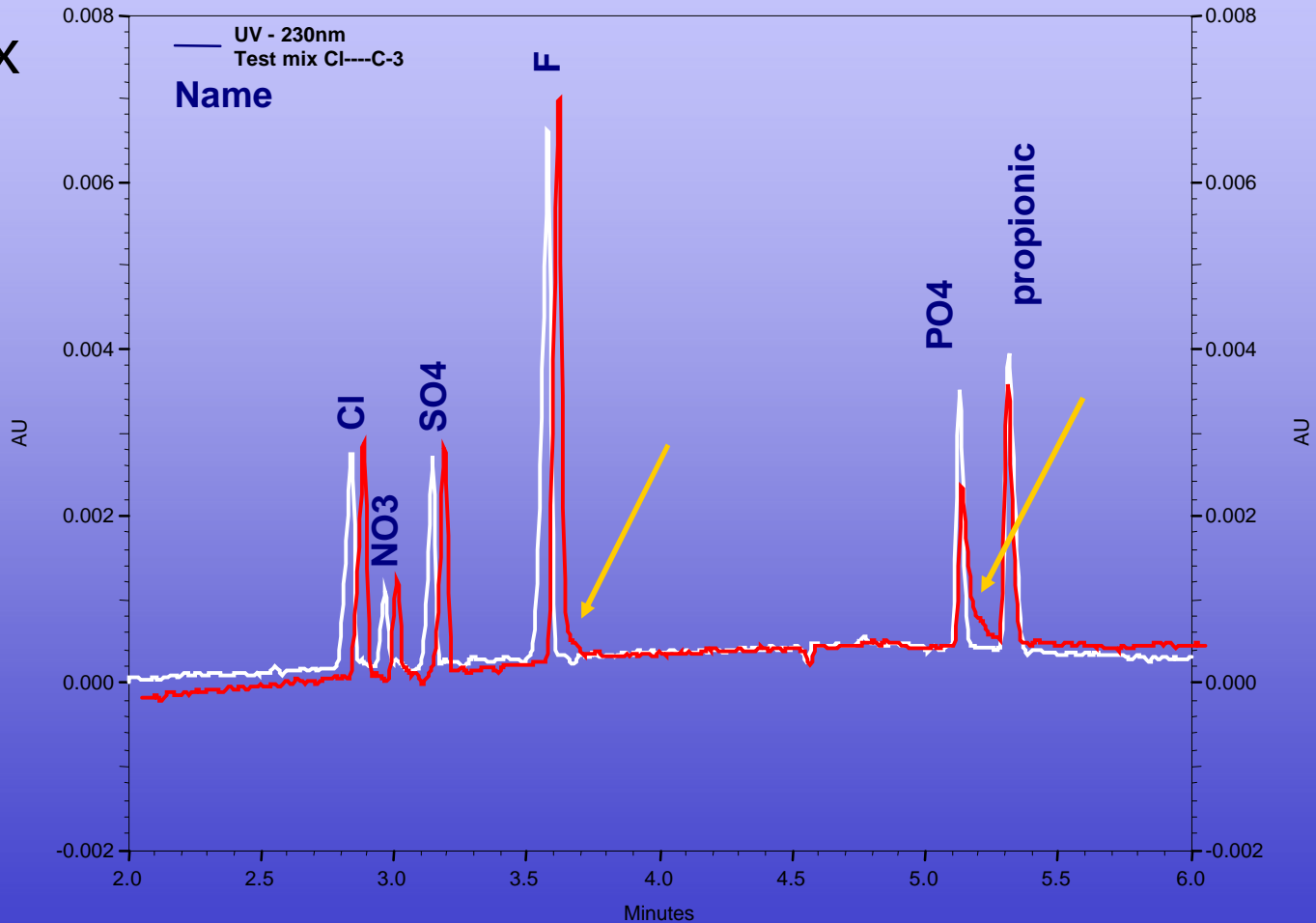
Analysis of Primaquine diphosphate (PQ)

- Sample: 200 mg/L PO_4
- Standards at 70%, 100% and 130%
- Octanoic Acid as I.S.
- Sequence: 3 x alternating standards and samples
- CEofix™ Anions5

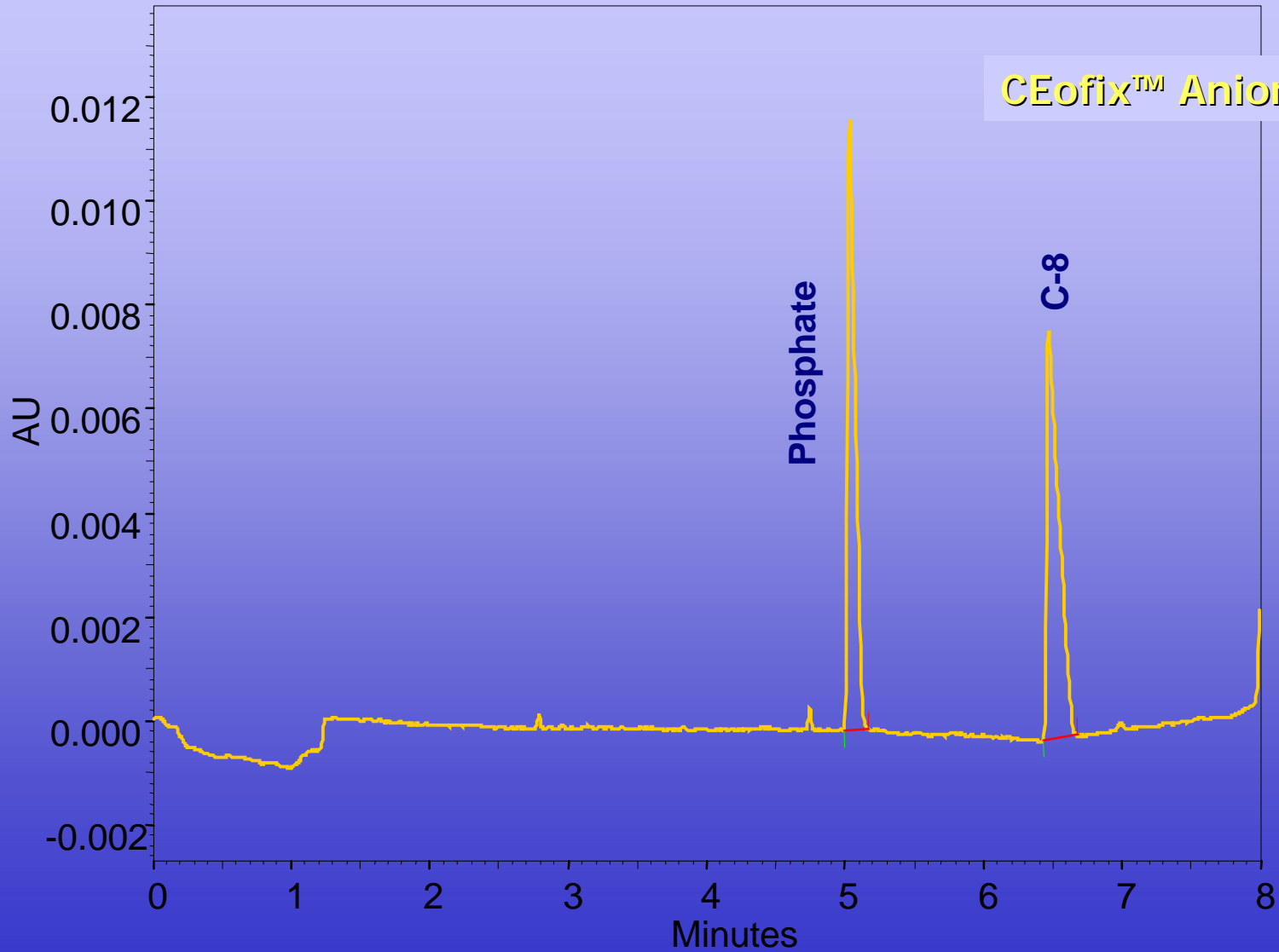
Problem of phosphate: peak tailing

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testmix

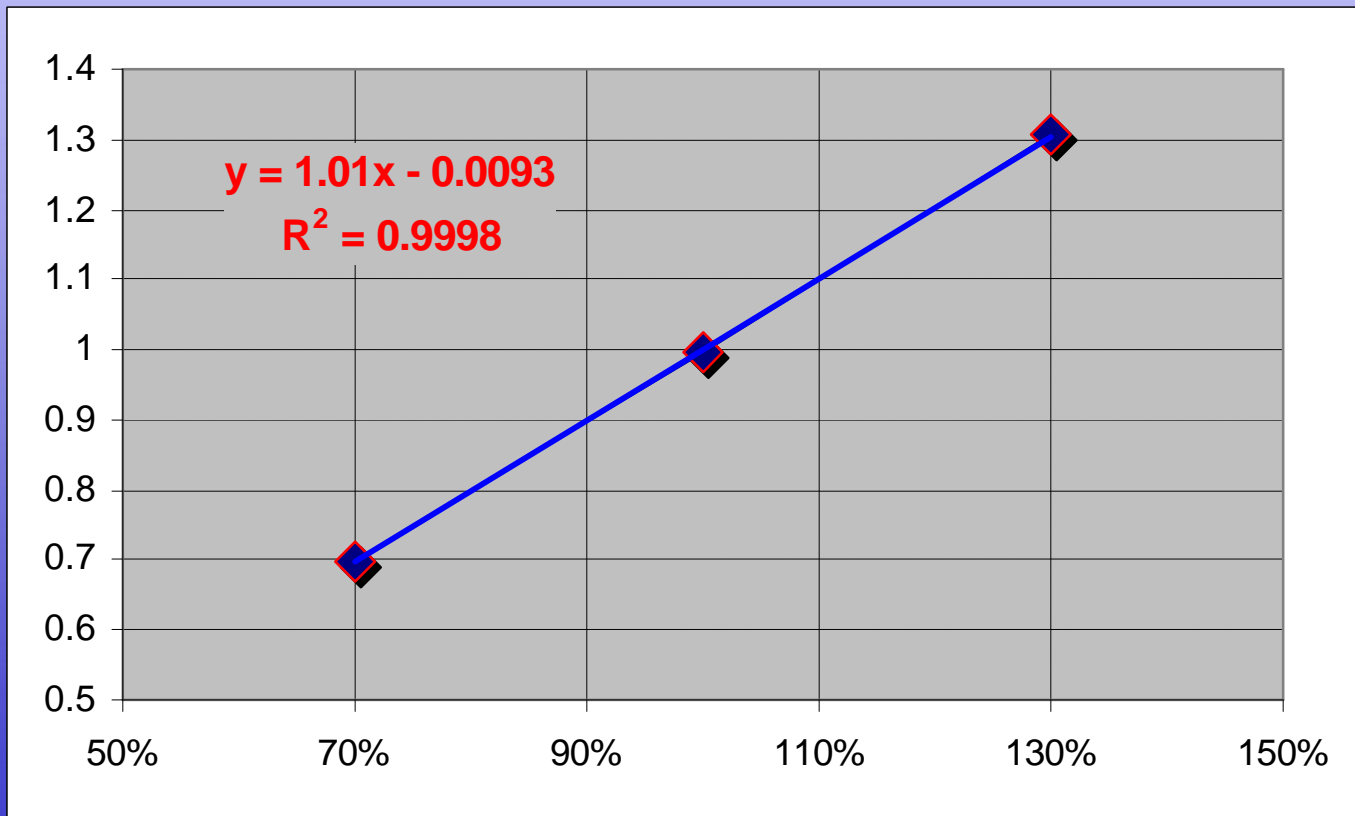


rinse H₂O ? rinse HCl 0.1M



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Linearity



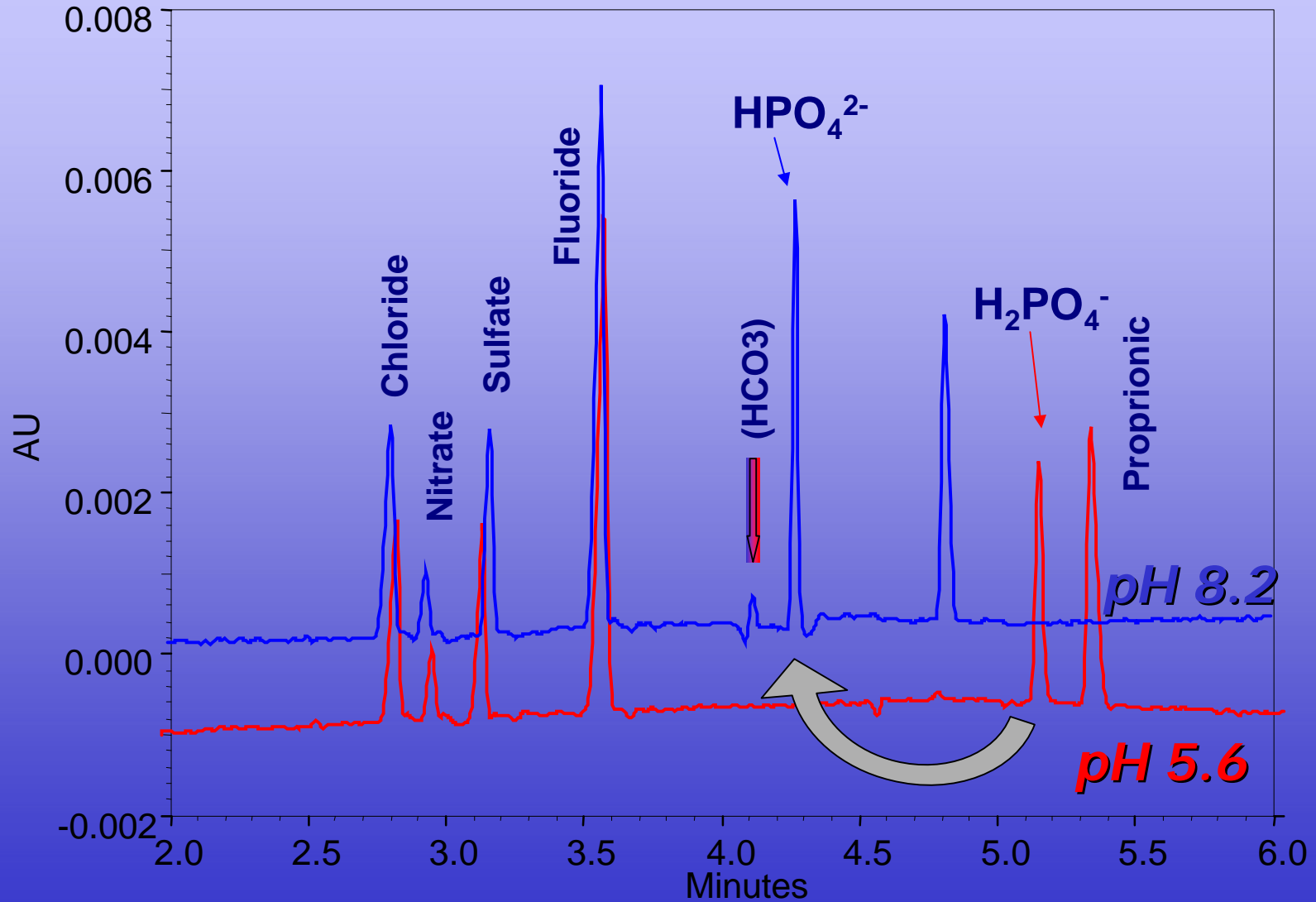
Results

	Migration Time		Area
	Phosphate	I.S.	I.S.
Mean	5.03	6.468	49749
%SD	0.25	0.21	2.79

	Sample PQ % Target Value
Run 4	99.82
Run 8	99.62
Run 12	99.81
Mean	99.75
%SD	0.11

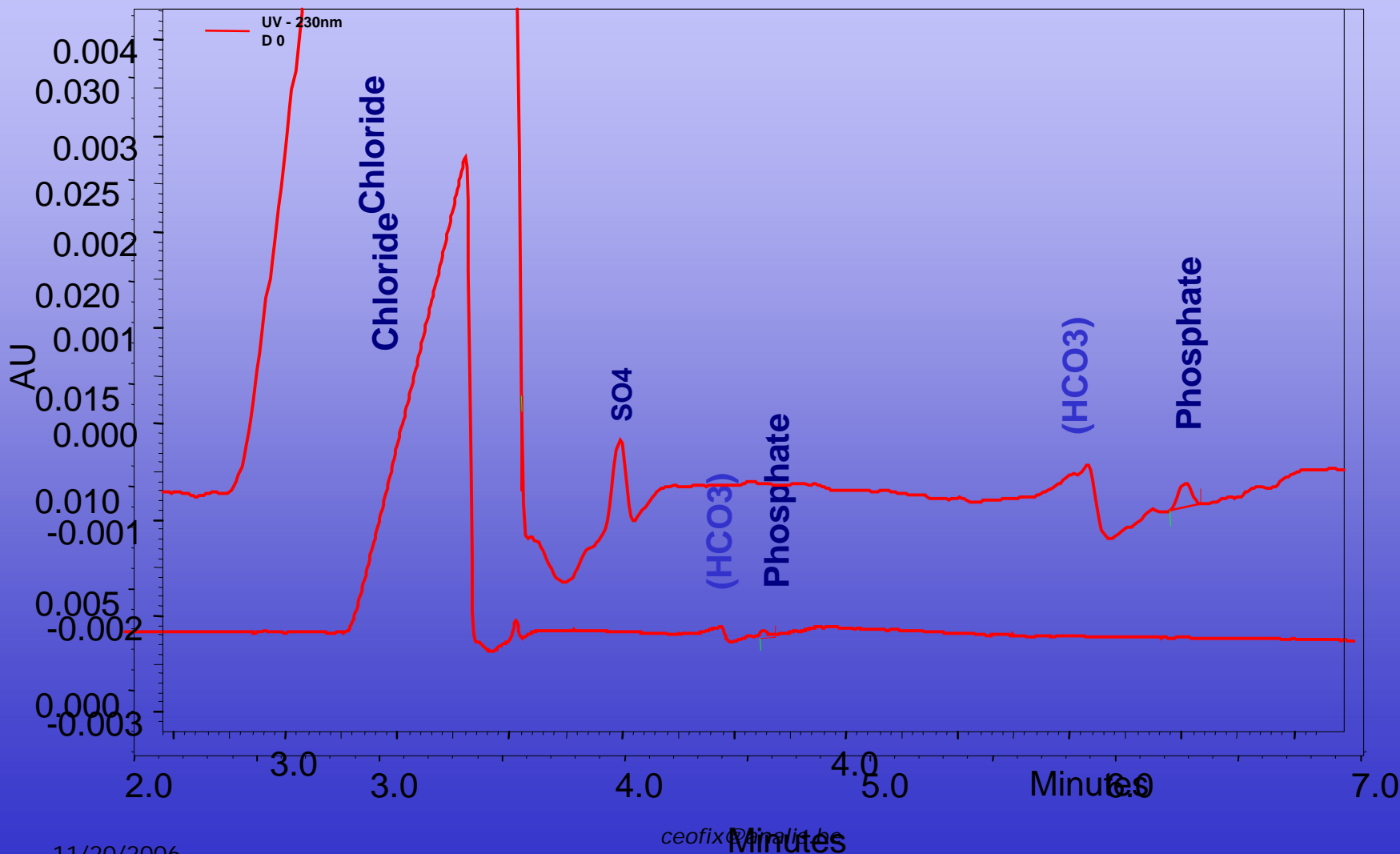
Impurity Analysis

- Phosphate impurity of a drug HCl salt
- Low level of impurity
- High level of Chloride
- CEofix™ Anions8



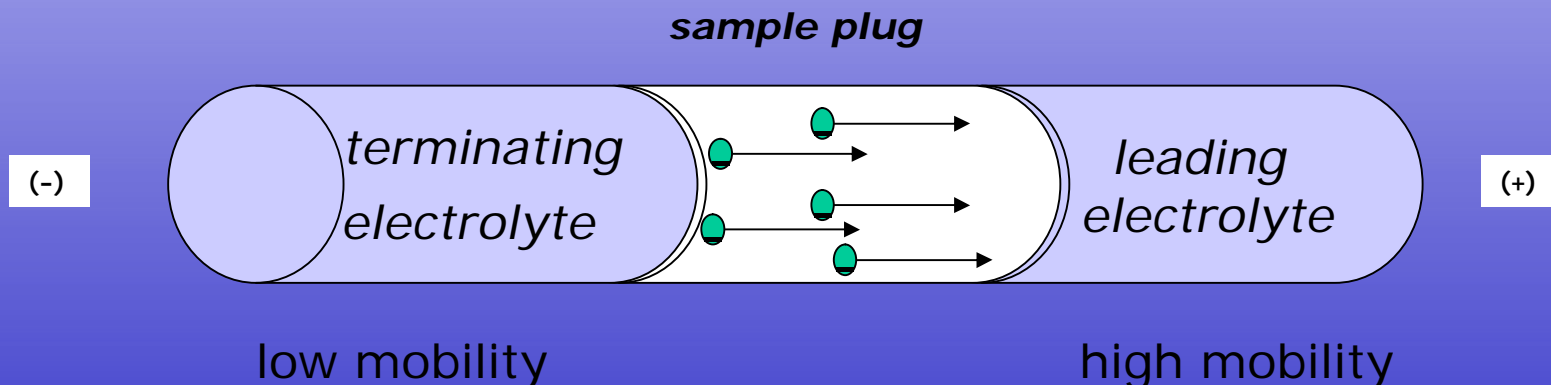
400 mg/L chloride and 0.1% phosphate

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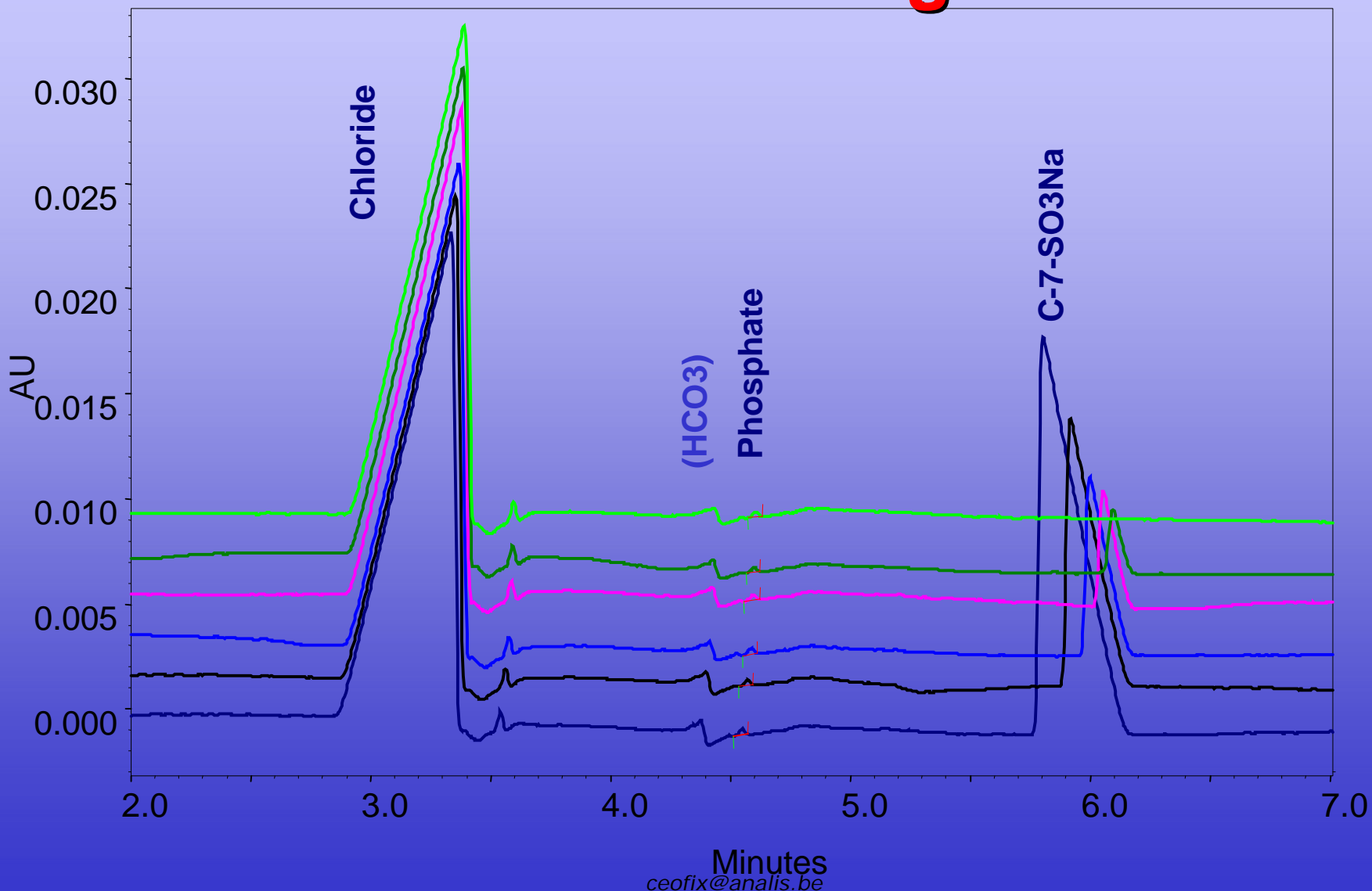


IsoTachoPhoretic Stacking (ITP)

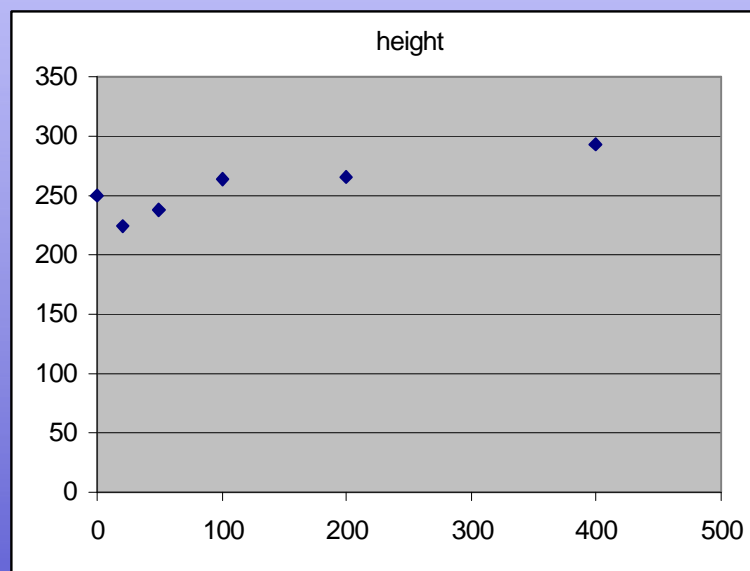
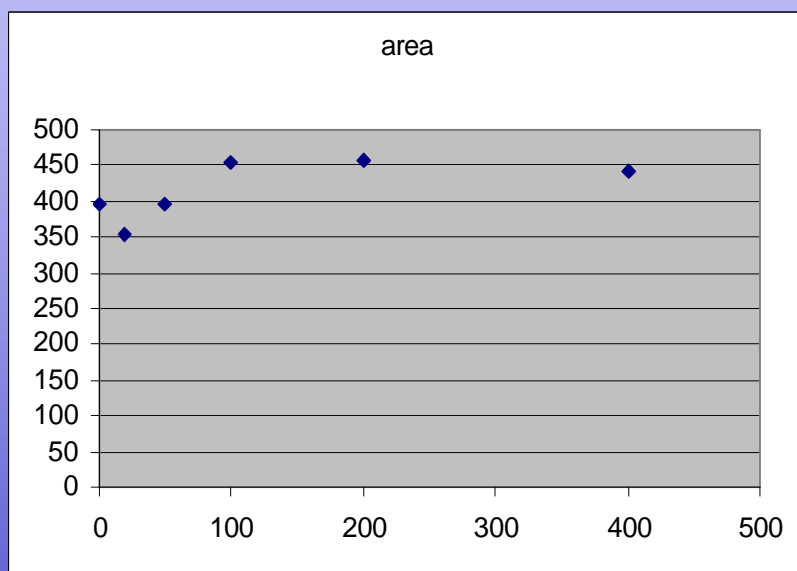
- Chloride high mobility ion -> de-stacking effect
- Addition of low mobility ion (in sample) -> re-stacking effect



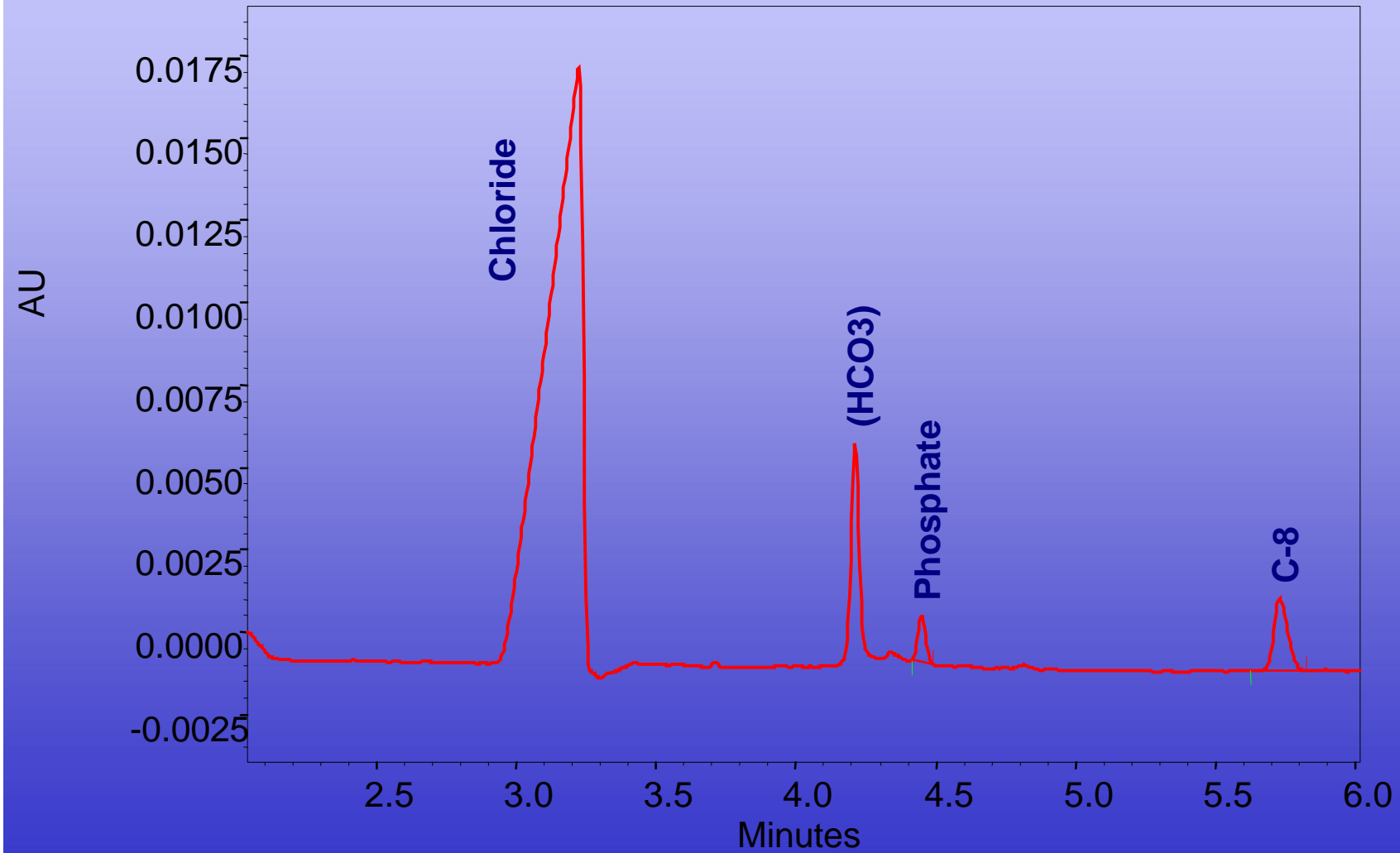
Heptane sulfonate as terminating ion



Effect of increasing amount of Heptanesulfonate as terminating ion



- SD on area: 9.82%
- no need for ITP re-stacking



Conclusion

- CE ion analysis with indirect UV detection is able to analyze
 - phosphate as counterion in a drug
 - phosphate as impurity in a drug
- Easy and fast method